Early Stages of Apomyoglobin Folding

Asghar Razavia, Heinrich Roderb, Vincent Voelza

^a Department of Chemistry, Temple University, Philadelphia, Pennsylvania 19122, United States b Fox Chase Cancer Center, Philadelphia, Pennsylvania 19111, United States

Introduction

Apomyoglobin (apoMb) folding pathways contain several intermediate states, identified by time-resolved fluorescence spectroscopy, which form on the microsecond to millisecond time scale. (Cite Xu and Roder JPCB) While structural information about these intermediates is extremely useful in understanding protein folding mechanisms, such states are short-lived and sparsely populated, making experimental characterization is a challenging task. To gain further insight, we turn to molecular simulations to model folding intermediates in atomic detail. The folding of apoMb follows a $U \rightarrow M \rightarrow N$ mechanism, where the M state is thought to be analogous to a highly-populated acid-denatured state at pH 4. Here, as a first step towards computational characterization of the M and N states, we perform large numbers of microsecond simulations at pH 4 and pH 7, restrained by experimentally determined H/D exchange protection factors. Our results are in good agreement with experimental observables, representing the first steps toward an atomically-detailed description of the conformational ensembles corresponding to the M and N states of apomyoglobin.

Objective

- Implement the H/D protection factors restraints in simulations.
- Characterize the N and M states at pH = 7and pH = 4.
- Build a kinetic network model to describe apomyoglobin folding at low and neutral

Methods and Simulations

Implementation of protection factor-restrained simulations in GROMACS

Maximum Entropy is the least biased way for enforcing restraints in simulations

$$U'(x) = U(x) + \alpha f(x)$$

$$U = U_0 + \frac{e^{-b(x-x0)}}{1+e^{-b(x-x0)}} \times k$$

$$k = \text{force constant } (\frac{kJ}{nm})$$

$$y = \frac{e^{-b(x-x0)}}{1+e^{-b(x-x0)}}$$

$$x0 = 6.5 \text{ Å}$$

$$b = 5.0$$

$$U(x) + \alpha f(x)$$

$$0.0$$

$$0.0$$

$$0.5$$

$$0.0$$

$$0.5$$

$$0.0$$

$$0.5$$

$$0.0$$

$$0.5$$

$$0.0$$

$$0.5$$

$$0.0$$

$$0.0$$

$$0.5$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

$$0.0$$

Distances between amide hydrogen and all oxygen atoms within 6.5Å of amide groups were restrained.

Simulations

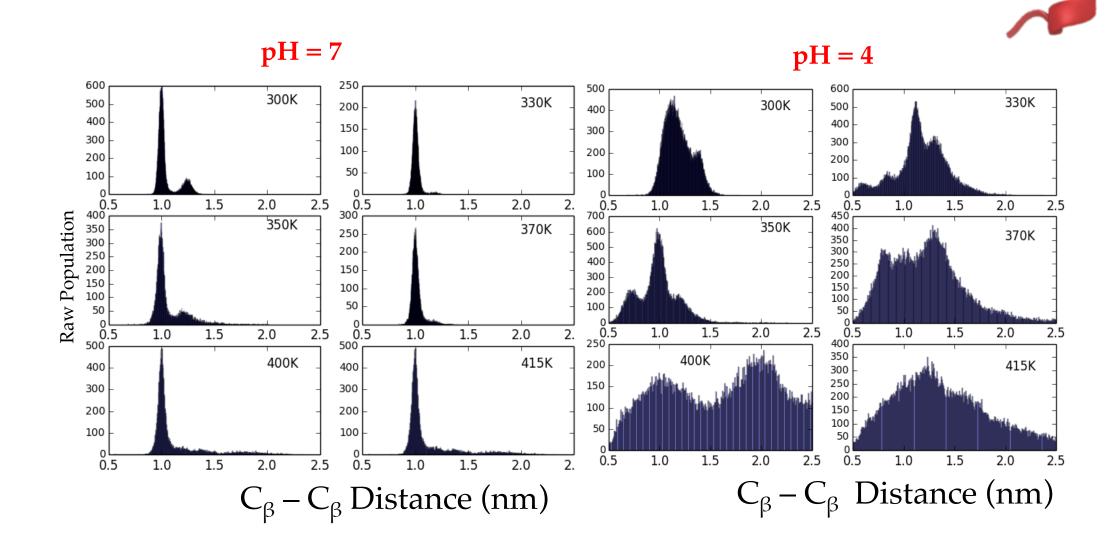
	Temperatures (K)	300, 330, 350, 370, 400, 415
pH = 4	Force Constants (kJ/nm)	0.0,0.1,0.2,0.3,0.4,0.5,0.6,0.7,0.8,0.9,1.0,1.1,1.2,1.5
TT P	Temperatures (K)	300, 330, 350, 370, 400, 415
pH = 7	Force Constants (kJ/nm)	0.0,0.5,0.7,1.0,1.2,1.5,2.0

Total simulation times

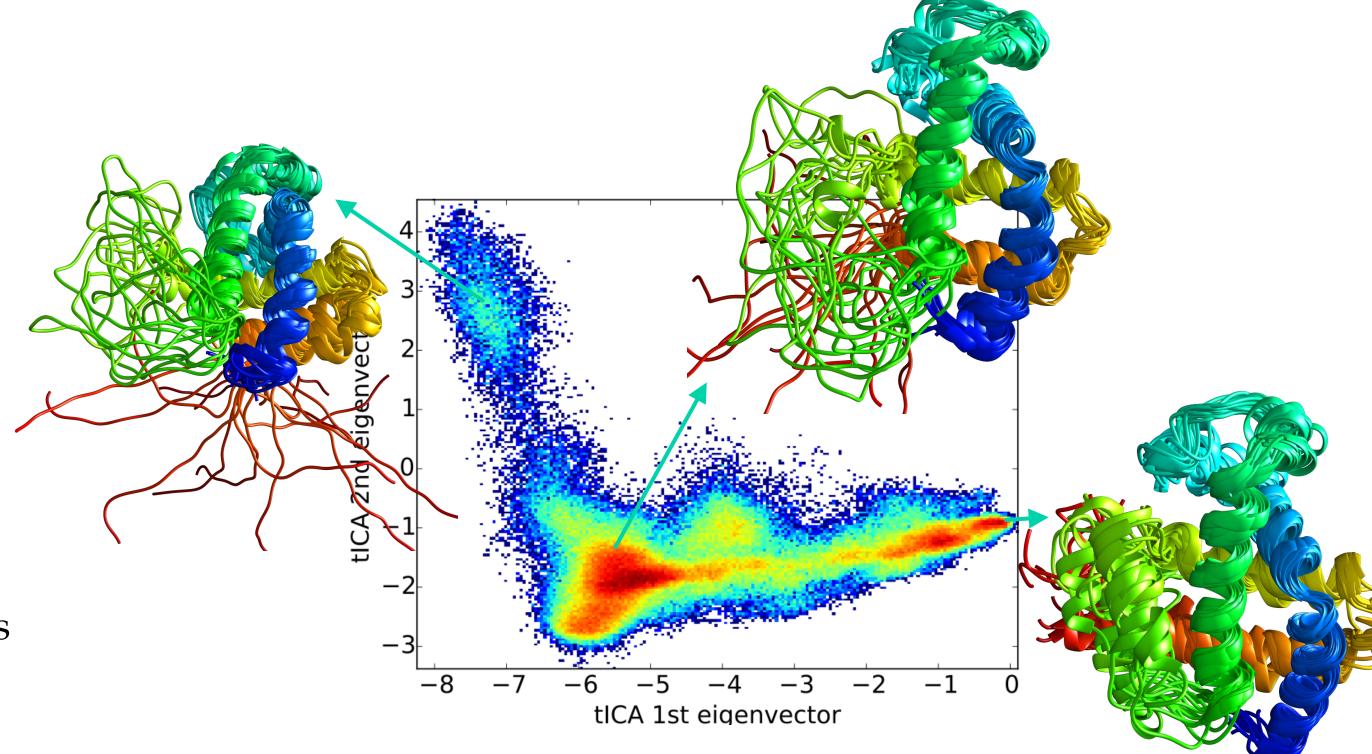
• pH=7: $\sim 100 \,\mu s$ • pH=4: $\sim 650 \,\mu s$

Results

His24 and His119 interaction is stable at pH = 7 even above melting temperature, but is disrupted at pH = 4.

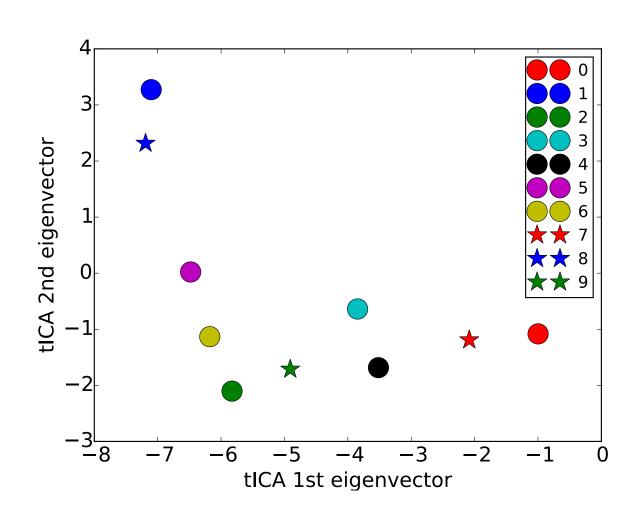


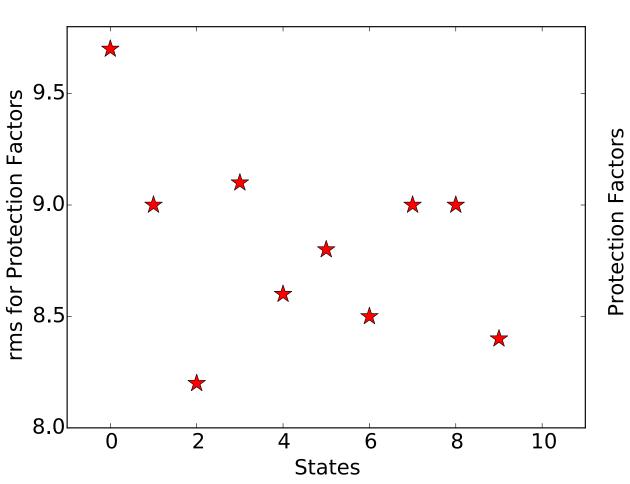
Free energy landscape of pH 7 restrained simulations contains several basins, which differ in degree of helicity for the F helix and C-terminal of the H helix.

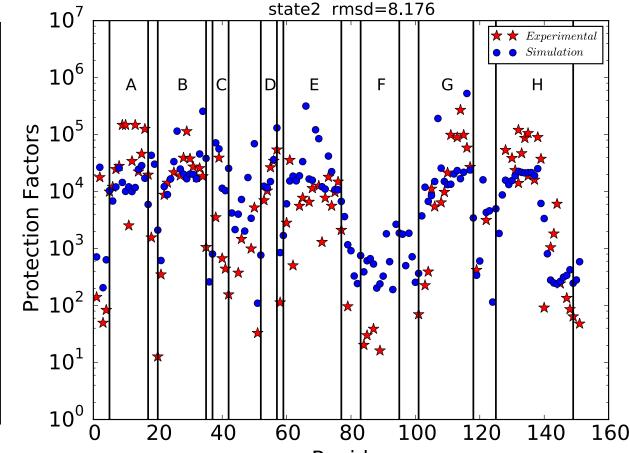


Results

Clustering the free energy landscape into 10 states and calculating the protection factors for each states, reveals state 2 as the most resembling of the apo state at neutral pH for myoglobin.







Conclusions

- At neutral pH, the His24-His119 interaction is highly stable, even as temperature increases to 415 K.
- At low pH (pH 4), the His24-His119 interaction is broken even at room temperature.
- We successfully implemented H/D restrains in our MD simulations and reproduced the experimental protection factors.

Future Direction

 Reweighting the restrained simulation data to obtain unbiased kinetic transitions for constructing Markov State Models of apoMB folding.

Acknowledgments

This research was supported in part by the National Science Foundation through major research instrumentation grant number CNS-09-58854.







• NSF MCB-1412508: Collaborative Research: Early Stages of Protein Folding Explored by Experimental and Computational Approaches

References

- B. Geierstanger, M. Jamin, B. F. Volkman, and R. L. Baldwin, *Biochemistry*, vol. 37, 4254-4265, 1998
- C. Nishimura, H. J. Dyson, and P. E. Wright, *PNAS*, vol. 102, 4765–4770, **2005**
- J. W. Pitera, and J. D. Chodera, *J. Chem. Theory Comput.* vol. 8, 3445–3451, **2012**
- M. Xu, O. Beresneva, R. Rosario, and H. Roder, J. Phys. Chem. B. vol. 116, 7014–7025, 2012



